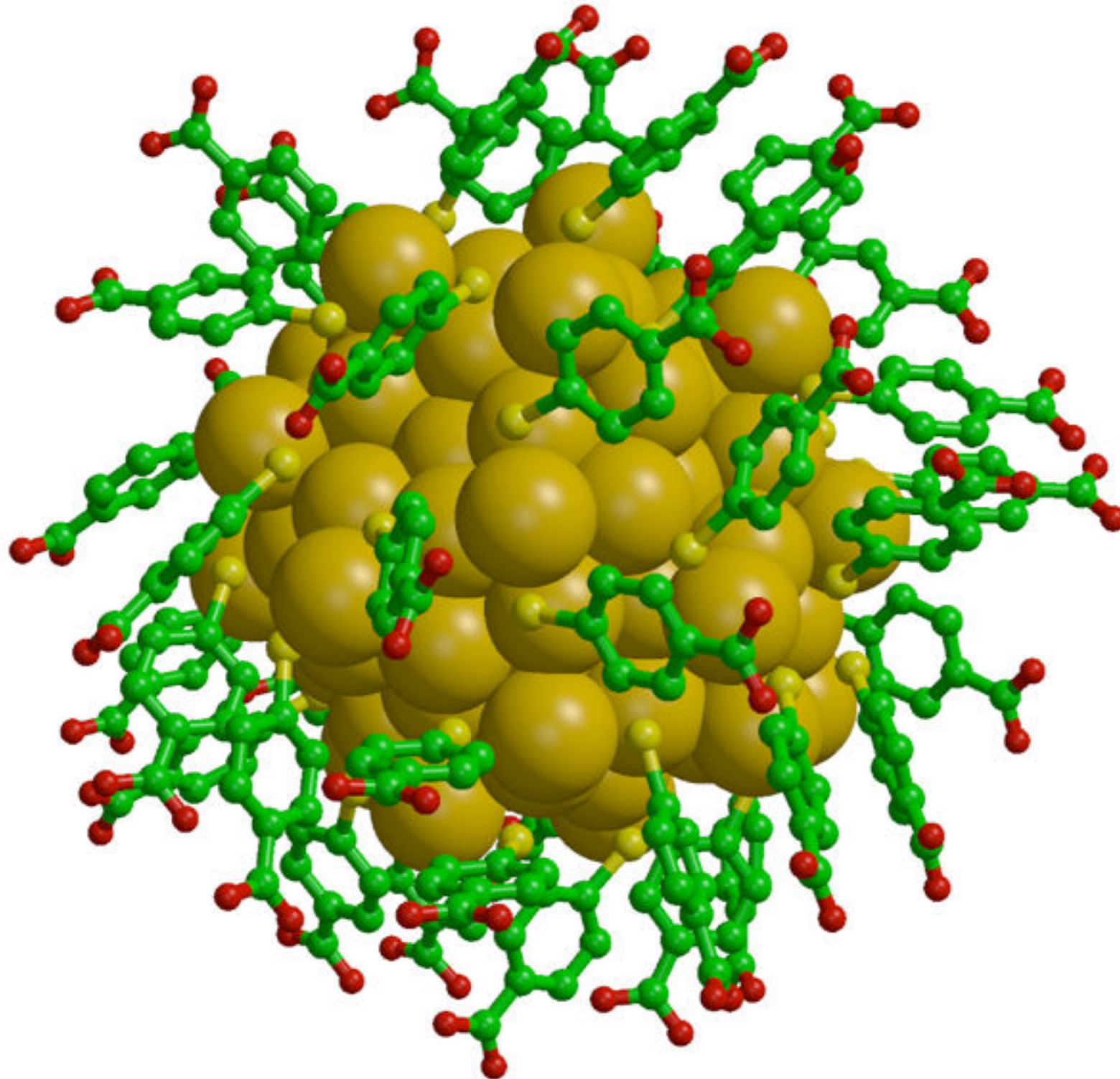


Bling Bling particles

Gerrit Groenhof

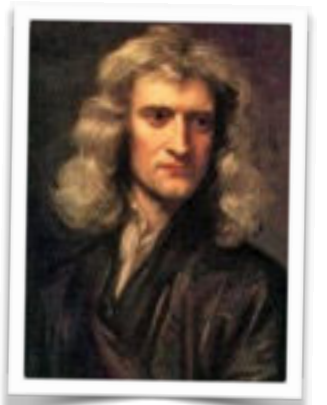
Nanoscience Center & Department of Chemistry



Molecular dynamics simulations

classical nuclei

$$\mathbf{F}_i = -\nabla_{\mathbf{R}_i} V(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) = m_i \frac{d^2}{dt^2} \mathbf{R}_i$$



Isaac Newton

Molecular dynamics simulations

classical nuclei

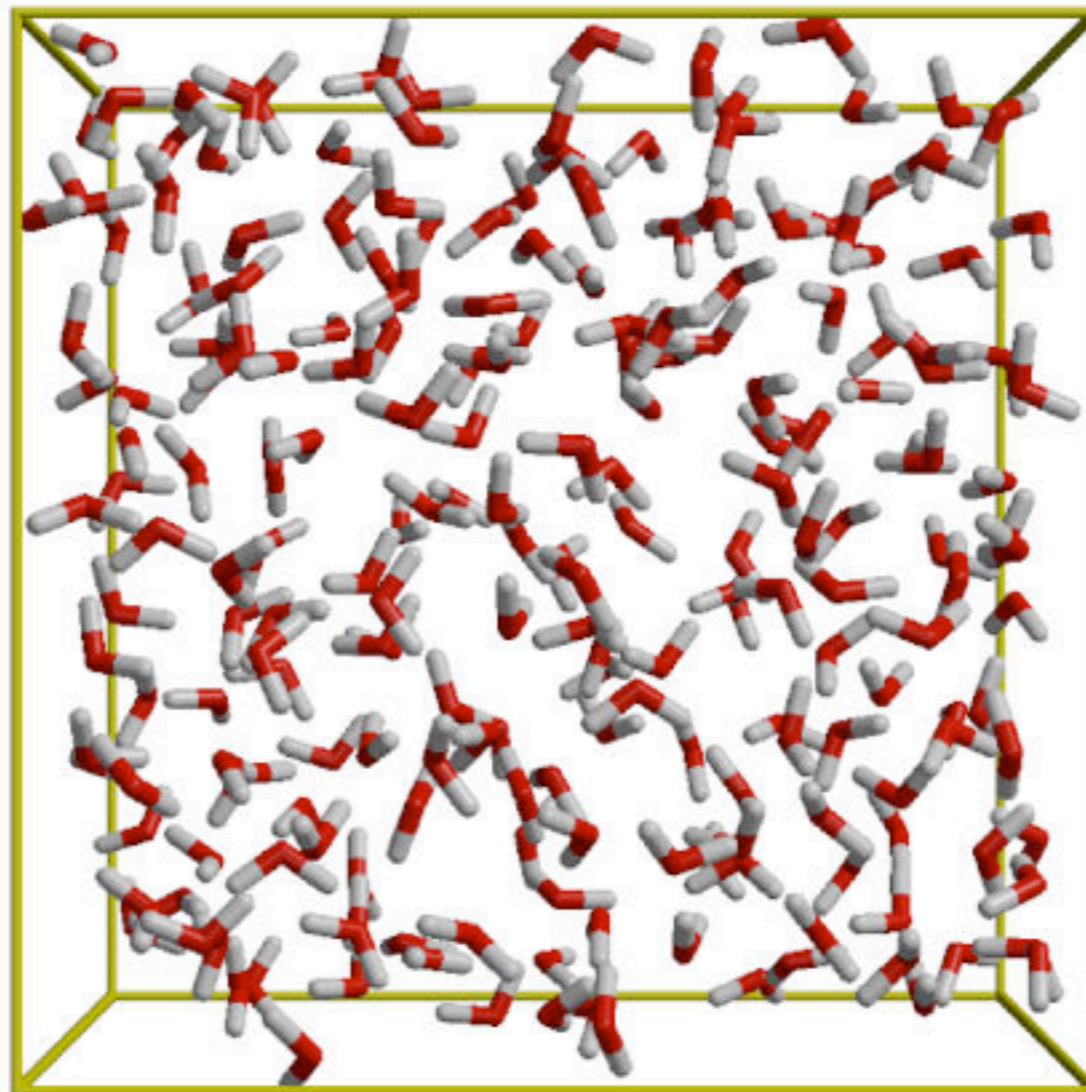
$$\mathbf{F}_i = -\nabla_{\mathbf{R}_i} V(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) = m_i \frac{d^2}{dt^2} \mathbf{R}_i$$

trajectory

jiggling & wiggling



Isaac Newton



Molecular dynamics simulations

classical nuclei

$$\mathbf{F}_i = -\nabla_{\mathbf{R}_i} V(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) = m_i \frac{d^2}{dt^2} \mathbf{R}_i$$

potential energy functions



Isaac Newton

Molecular dynamics simulations

classical nuclei

$$\mathbf{F}_i = -\nabla_{\mathbf{R}_i} V(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) = m_i \frac{d^2}{dt^2} \mathbf{R}_i$$

potential energy functions

quantum chemistry

$$V^{\text{QM}}(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) =$$

$$\langle \Psi_e(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_n) | \hat{H}_e(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) | \Psi_e(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_n) \rangle$$



Isaac Newton



Erwin Schrödinger

Molecular dynamics simulations

classical nuclei

$$\mathbf{F}_i = -\nabla_{\mathbf{R}_i} V(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) = m_i \frac{d^2}{dt^2} \mathbf{R}_i$$

potential energy functions

quantum chemistry (QM)

$$V^{\text{QM}}(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) =$$

$$\langle \Psi_e(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_n) | \hat{H}_e(\mathbf{R}_1, \mathbf{R}_2, \dots, \mathbf{R}_N) | \Psi_e(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_n) \rangle$$

molecular mechanics (MM)

empirical functions with parameters

$$V^{\text{MM}}(R_1, R_2, \dots, R_N) = \sum_k v_k(R_i, R_j, R_k, R_l; \{p_k\})$$



Isaac Newton



Erwin Schrödinger

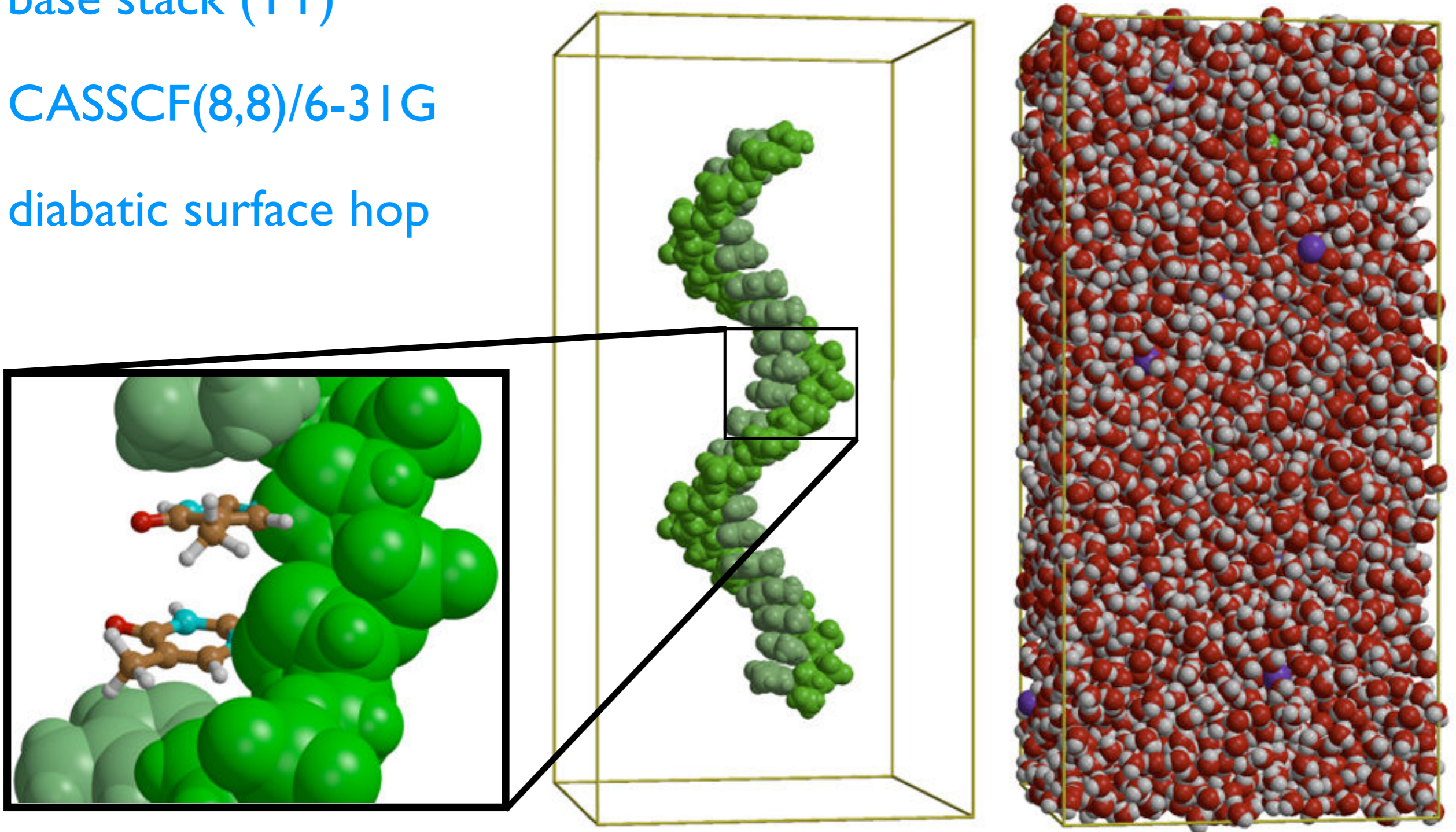
Observe while it all happens

computer simulations of DNA photodamage

base stack (TT)

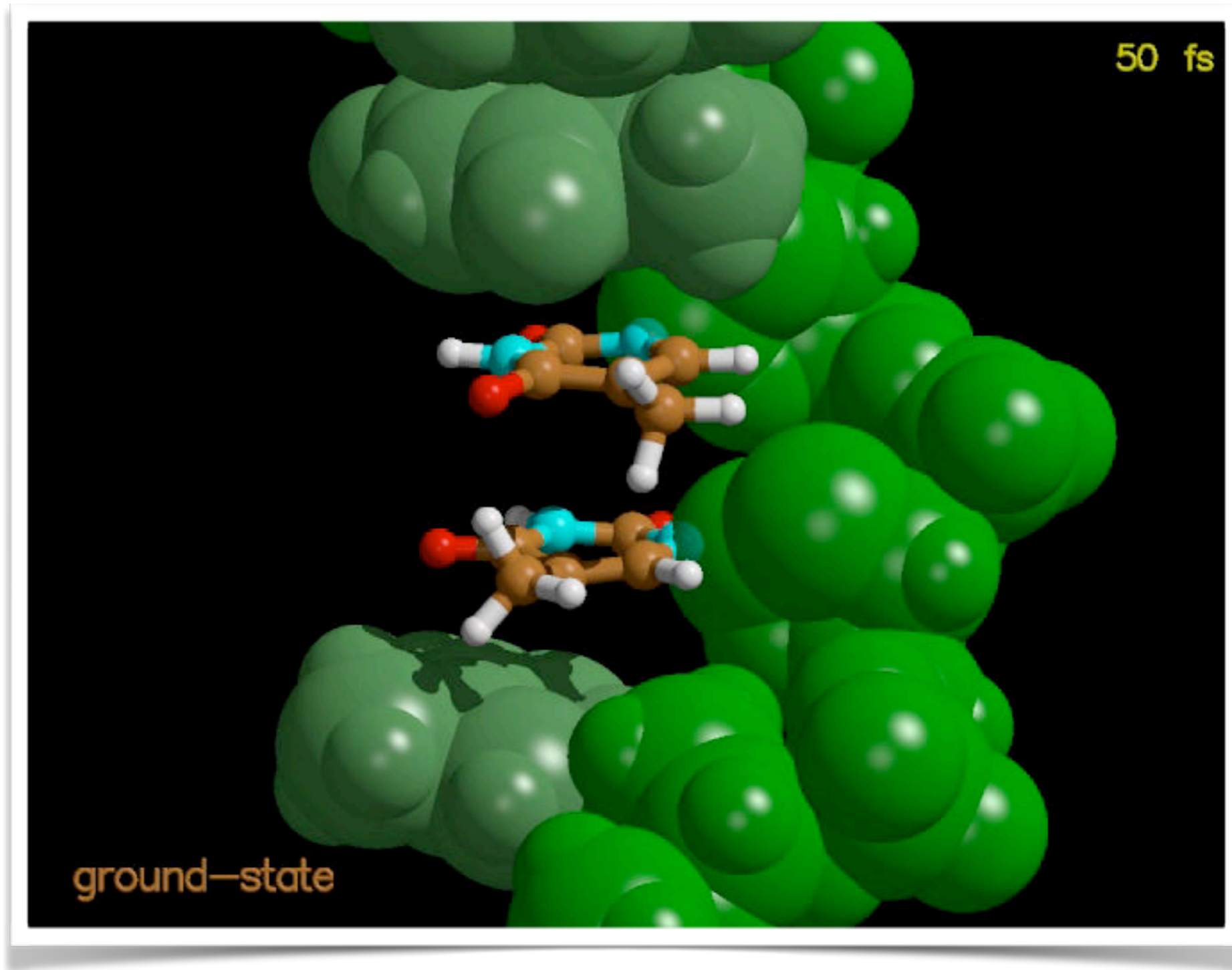
CASSCF(8,8)/6-31G

diabatic surface hop



Observe while it all happens

excited-state simulation of TT base stack



Today's project

Monolayer protected gold nanoparticles

claim: these are very important

who knows?

easy to publish

pK_a of nanoparticle?

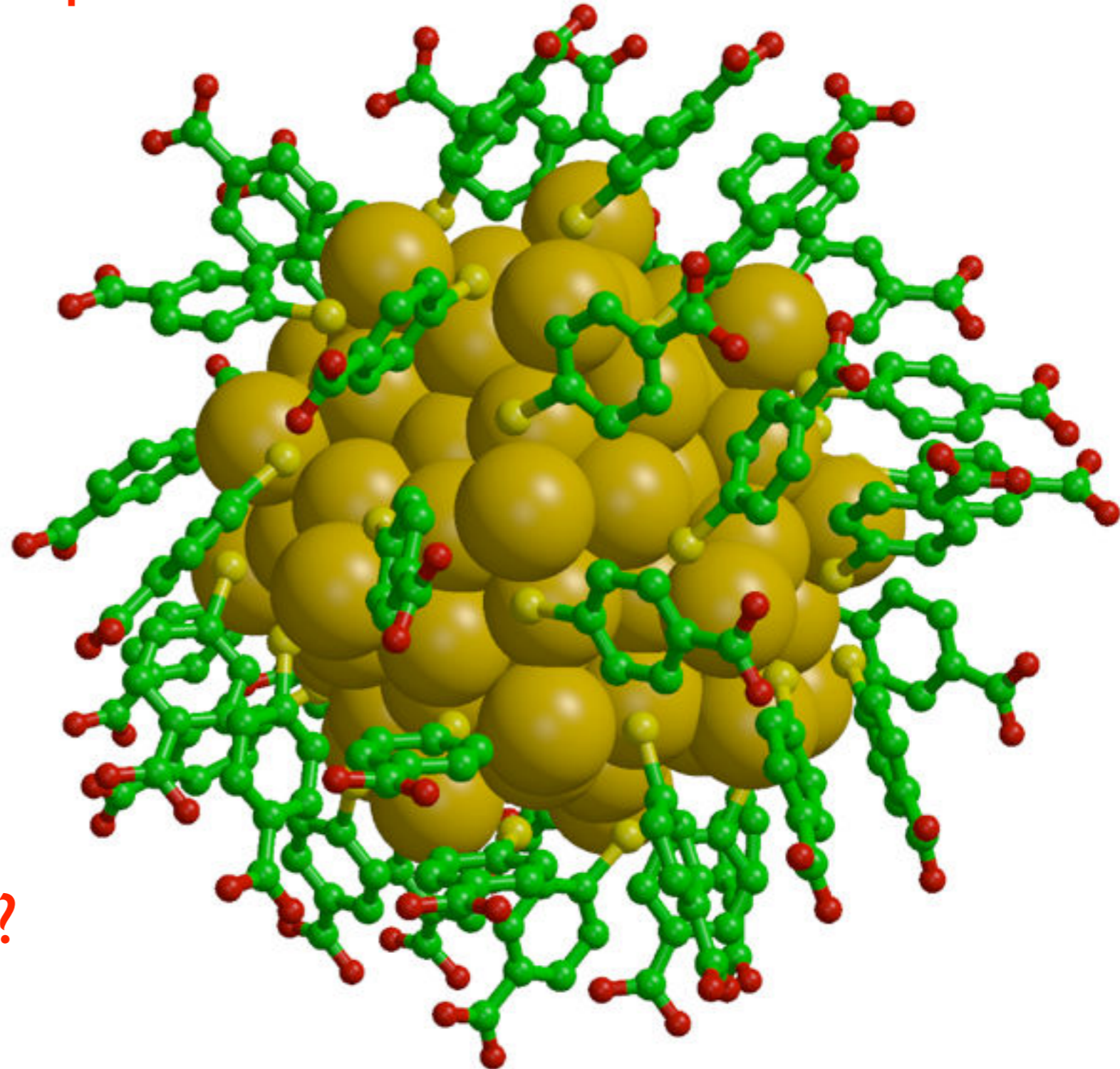
44 protonatable sites

anti-cooperativity

microscopic pK_a 's

What's actually a pK_a ?

who knows?



Constant pH molecular dynamics simulations

requirements

sample of configuration & protonation state space

control rate of transition between protonation states

fully atomistic description

explicit solvent



Serena
Donnini



Florian
Tegeler



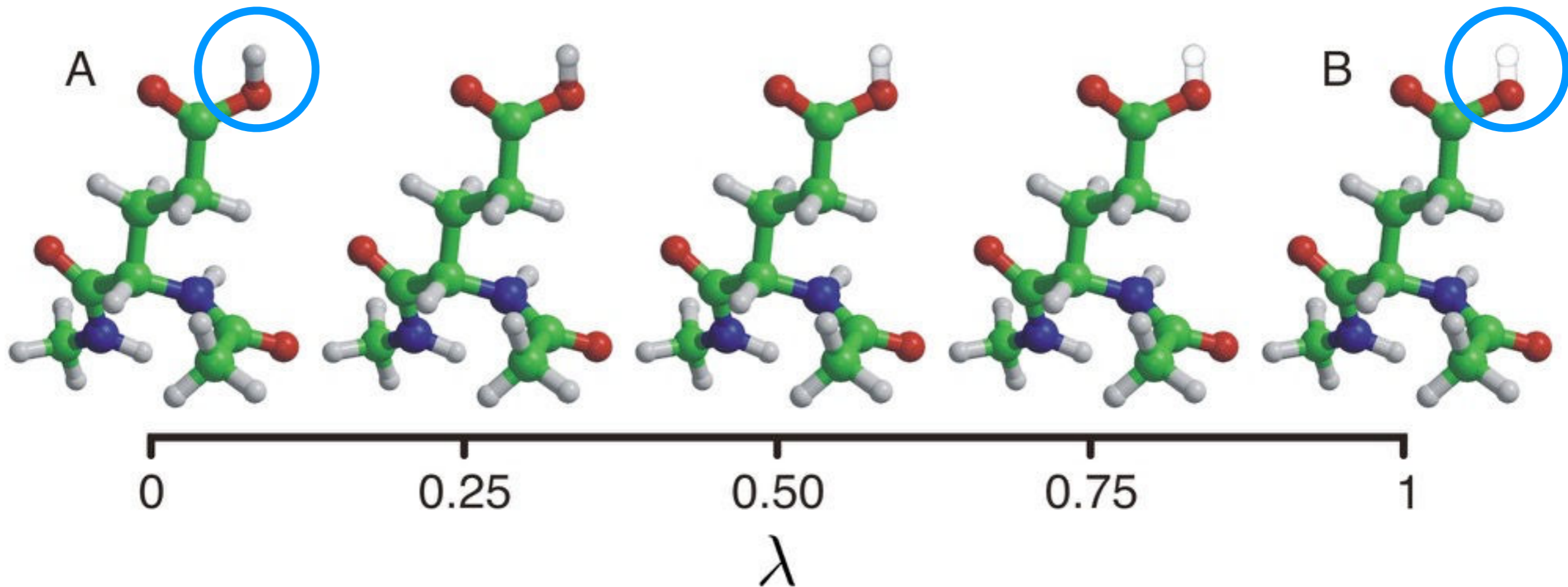
Helmut
Grubmüller



Donnini et al. *J. Chem. Theory. Comp.* 7 (2011) 1962

Molecular dynamics at constant pH

protons as extra degrees of freedom



dynamics of λ -particle (protonation)

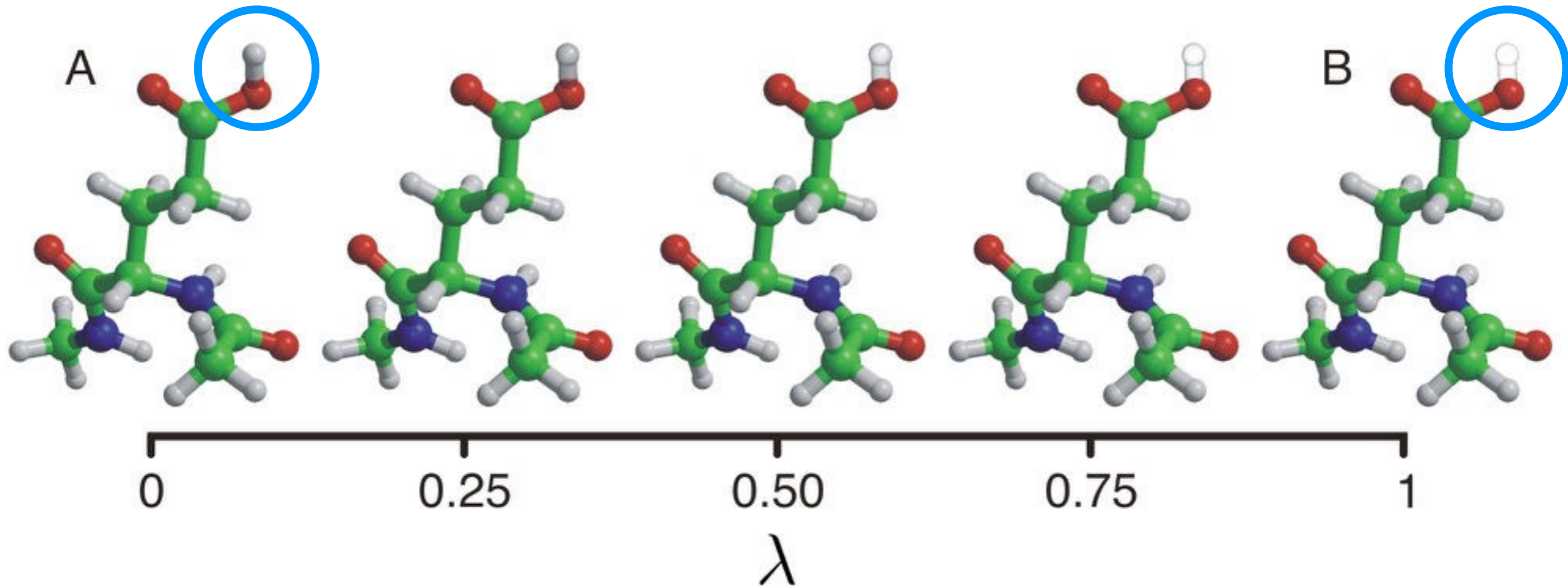
$$m_{\lambda} d^2 \lambda / dt^2 = -\partial V(\mathbf{x}, \lambda) / \partial \lambda$$

X. Kong & C.L. Brooks J. Chem. Phys. 105 (1996) 2414

M.S. Lee, J.F.R. Salsbury, C.L. Brooks Proteins 56 (2004) 738

Molecular dynamics at constant pH

protons as extra degrees of freedom



dynamics of λ -particle (protonation)

$$m_{\lambda} d^2 \lambda / dt^2 = -\partial V(\mathbf{x}, \lambda) / \partial \lambda$$

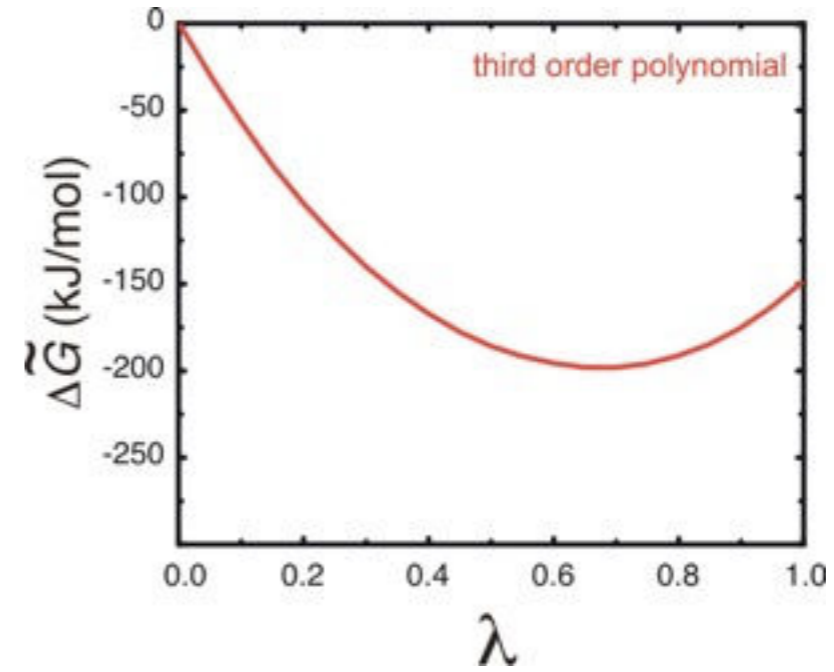
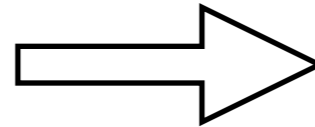
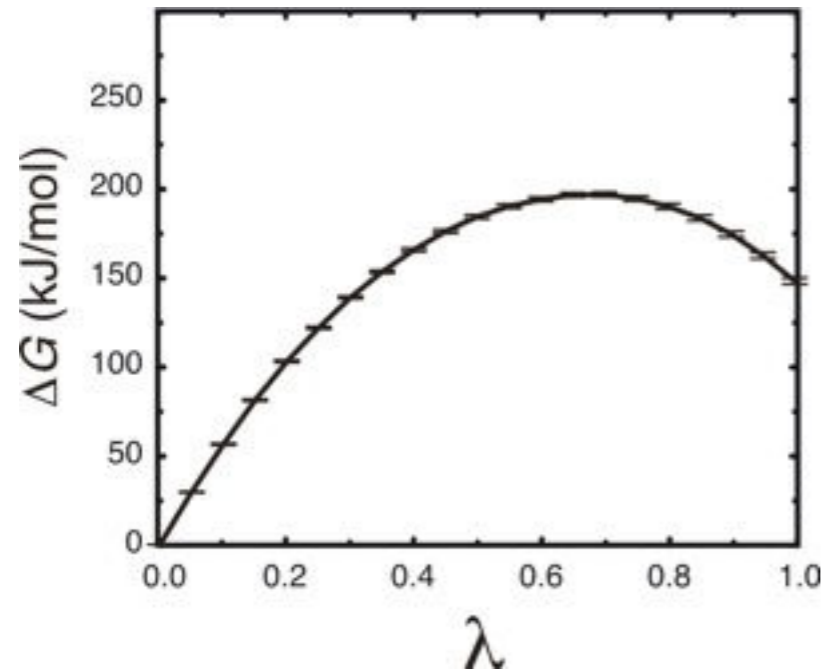
$$V(\mathbf{x}, \lambda) = (1 - \lambda)V^A(\mathbf{x}) + \lambda V^B(\mathbf{x}) + U(\lambda) +$$

$$\lambda RT \ln(10) [pK_{a, \text{ref}}^{\text{exp}} - pH] + \Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$$

$$V(\mathbf{x}, \lambda) = (1 - \lambda)V^A(\mathbf{x}) + \lambda V^B(\mathbf{x}) + U(\lambda) +$$

$$\lambda RT \ln(10) [pK_{a, \text{ref}}^{\text{exp}} - pH] + \Delta\tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$$

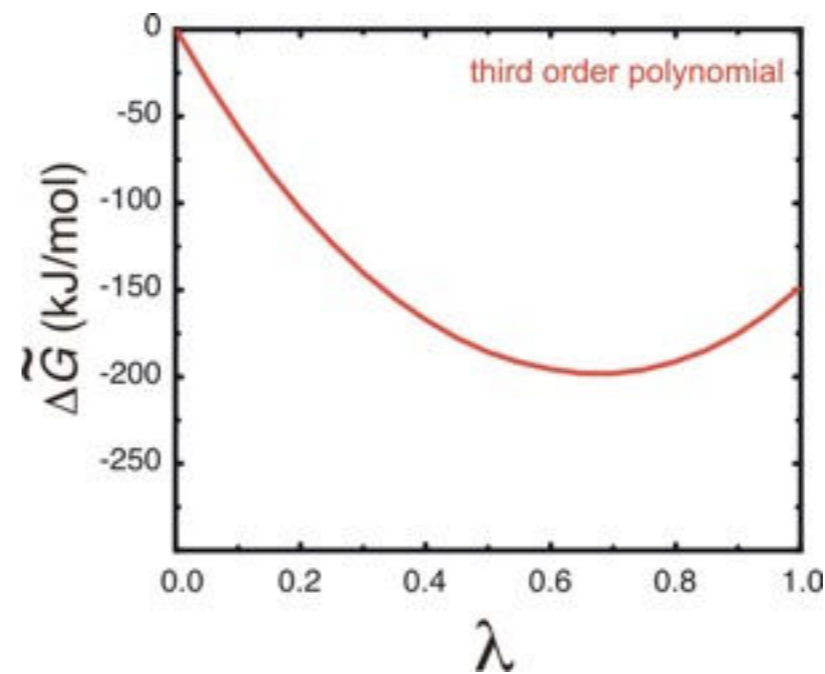
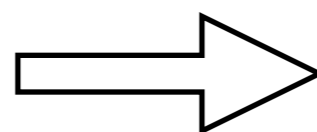
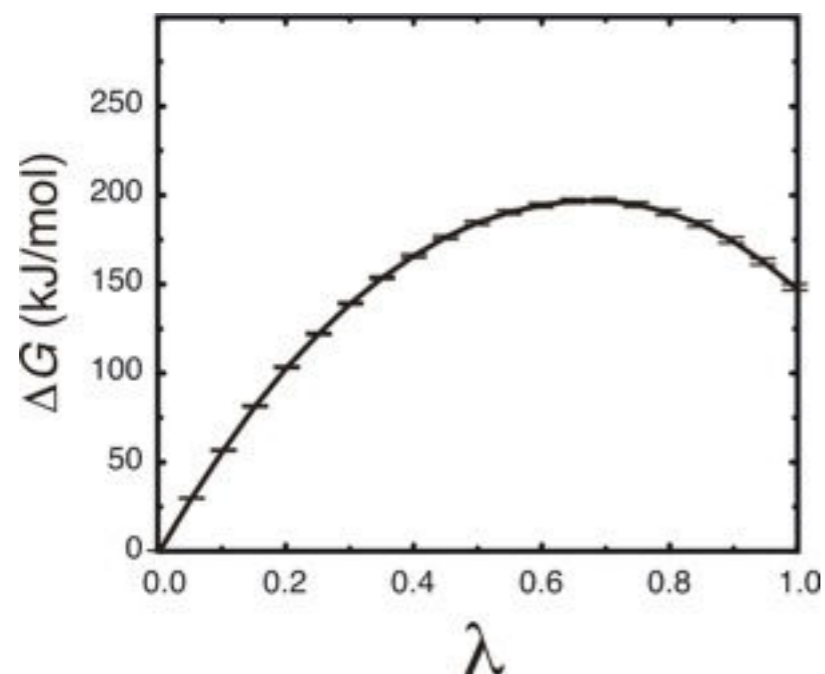
$\Delta\tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$ obtained by thermodynamic integration ($pH = pK_a$)



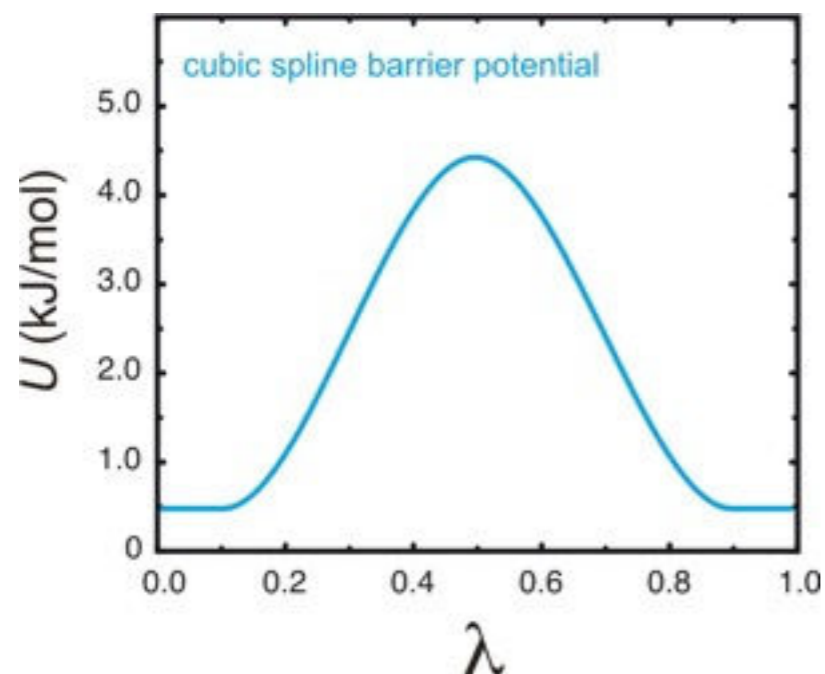
$$V(\mathbf{x}, \lambda) = (1 - \lambda)V^A(\mathbf{x}) + \lambda V^B(\mathbf{x}) + U(\lambda) +$$

$$\lambda RT \ln(10) [pK_{a, \text{ref}}^{\text{exp}} - pH] + \Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$$

$\Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$ obtained by thermodynamic integration ($pH = pK_a$)



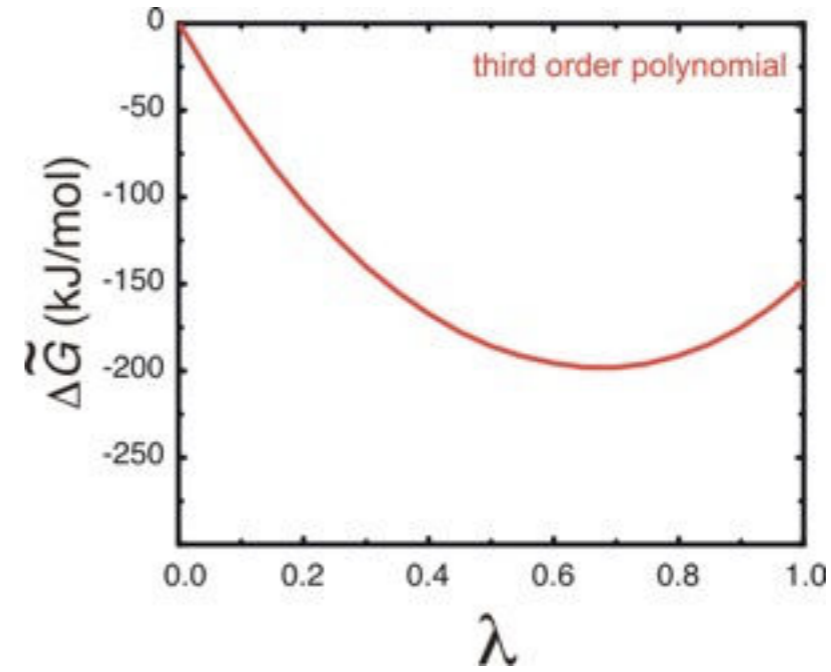
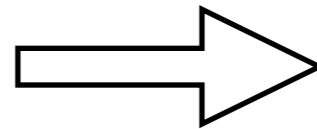
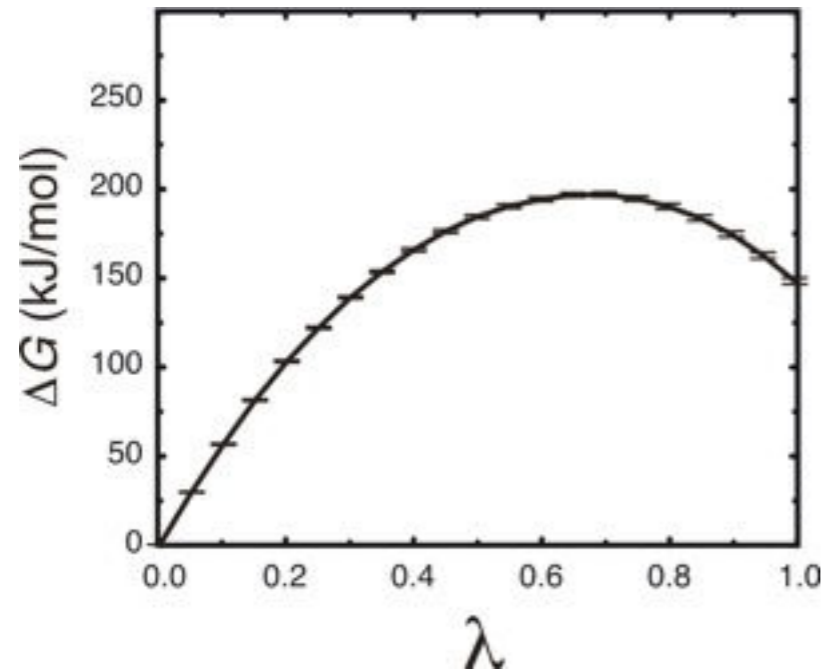
barrier potential



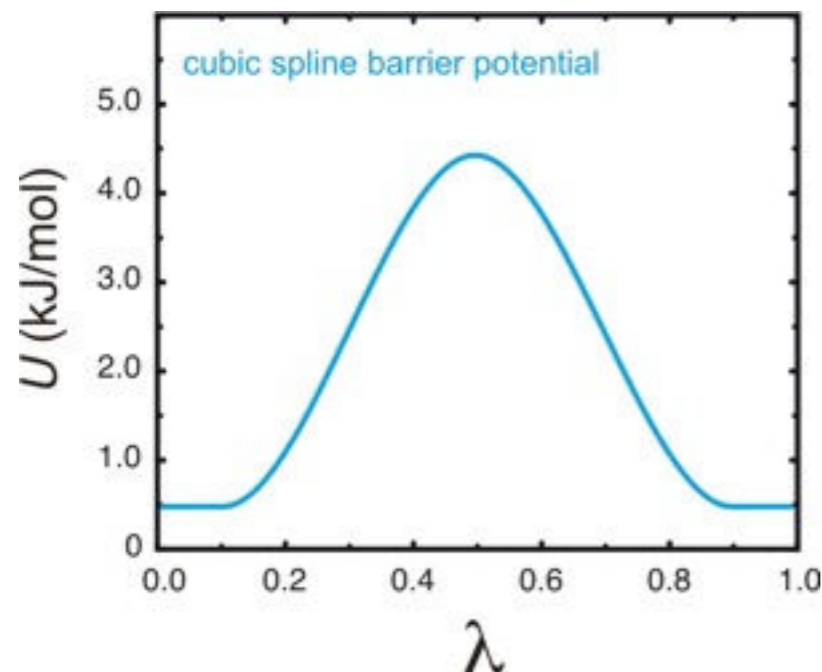
$$V(\mathbf{x}, \lambda) = (1 - \lambda)V^A(\mathbf{x}) + \lambda V^B(\mathbf{x}) + U(\lambda) +$$

$$\lambda RT \ln(10) [pK_{a, \text{ref}}^{\text{exp}} - \text{pH}] + \Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$$

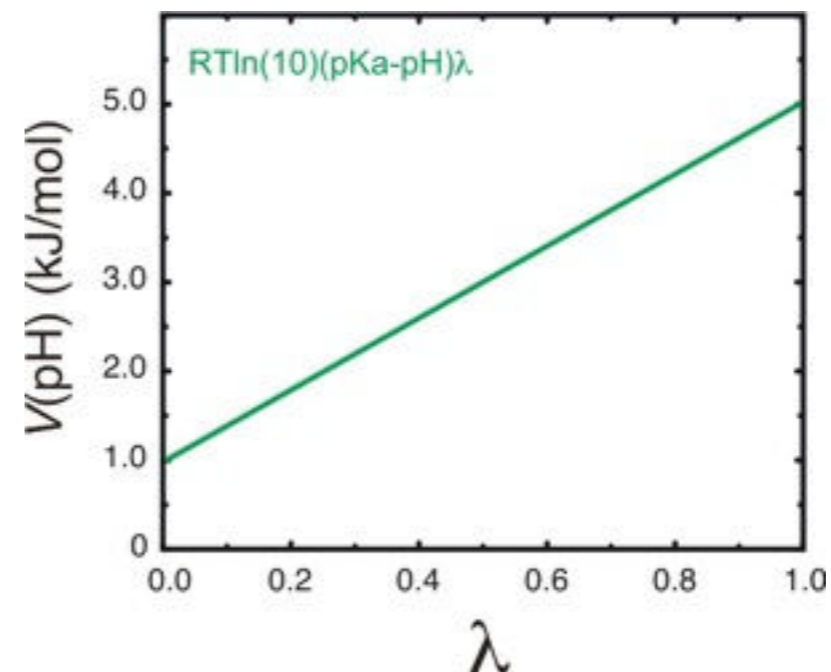
$\Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda)$ obtained by thermodynamic integration ($\text{pH} = \text{p}K_a$)



barrier potential

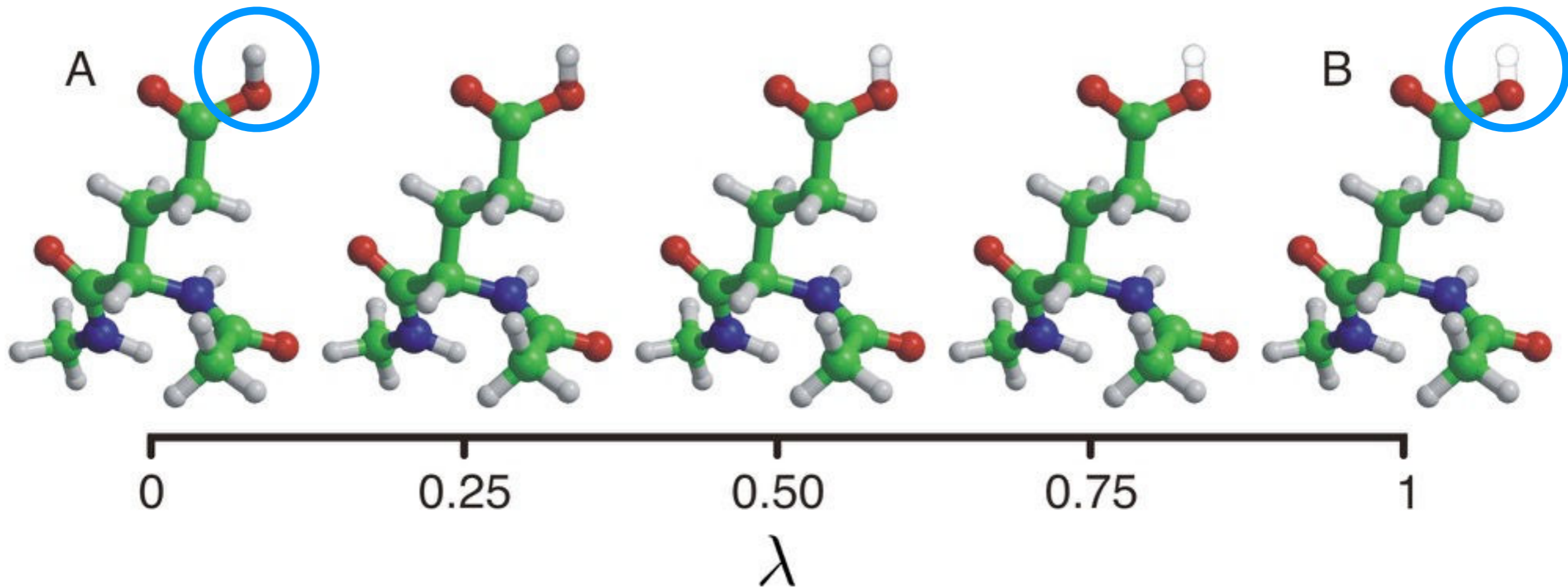


pH dependence



Molecular dynamics at constant pH

protons as extra degrees of freedom



dynamics of multiple λ -particles (protonation states)

$$m_{\lambda_i} d^2 \lambda_i / dt^2 = -\partial V(\mathbf{x}, \lambda_i) / \partial \lambda_i$$

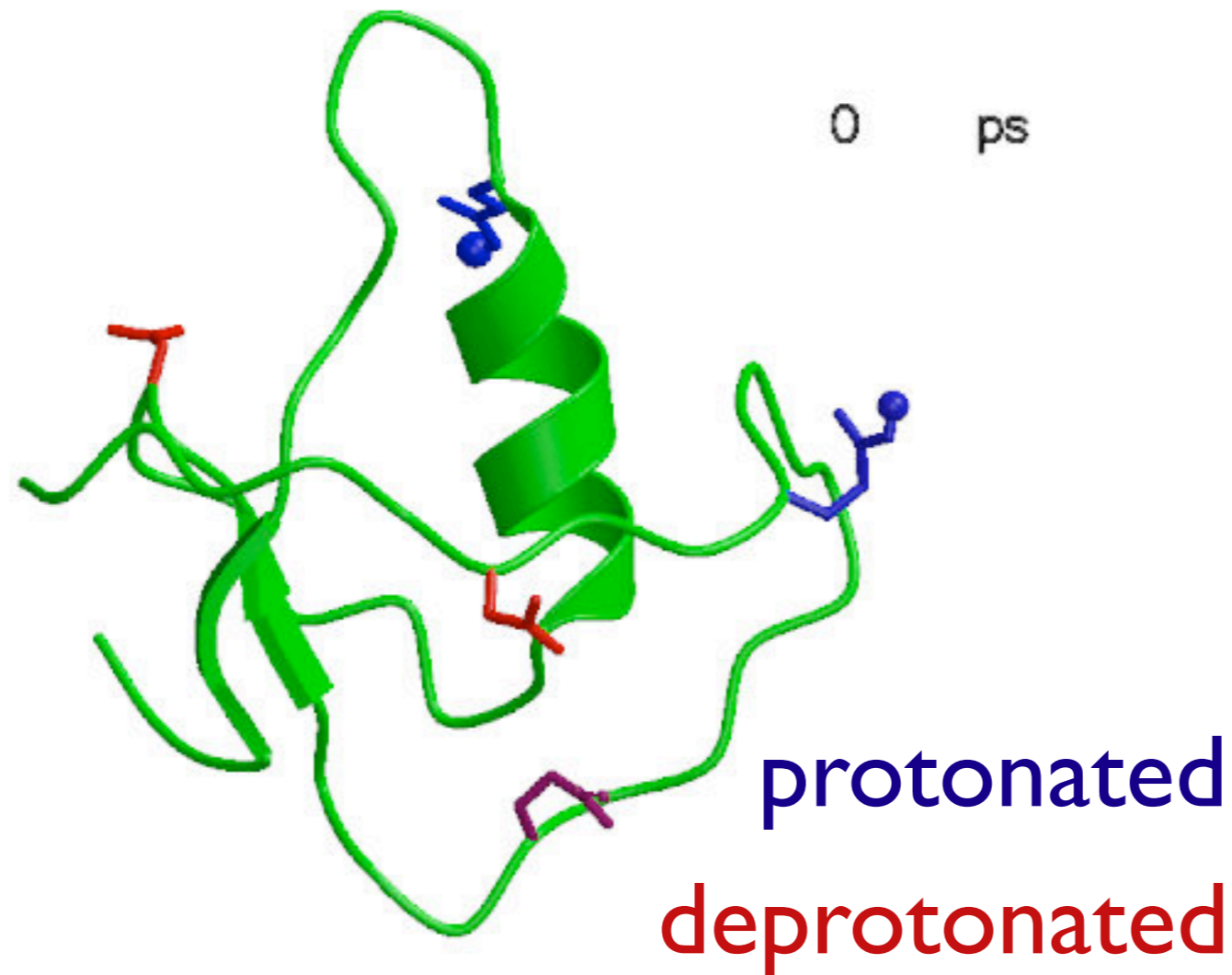
$$V(\mathbf{x}, \lambda_i) = (1 - \lambda_i) V^A(\mathbf{x}, \boldsymbol{\lambda}) + \lambda_i V^B(\mathbf{x}, \boldsymbol{\lambda}) + U(\lambda_i) +$$

$$\lambda_i RT \ln(10) [pK_{a, \text{ref}}^{\text{exp}} - pH] + \Delta \tilde{G}_{\text{MM}}^{\text{corr}}(\lambda_i, \boldsymbol{\lambda})$$

Molecular dynamics at constant pH

time-dependent protonation states

third domain of turkey ovomucoid inhibitor at pH = 4



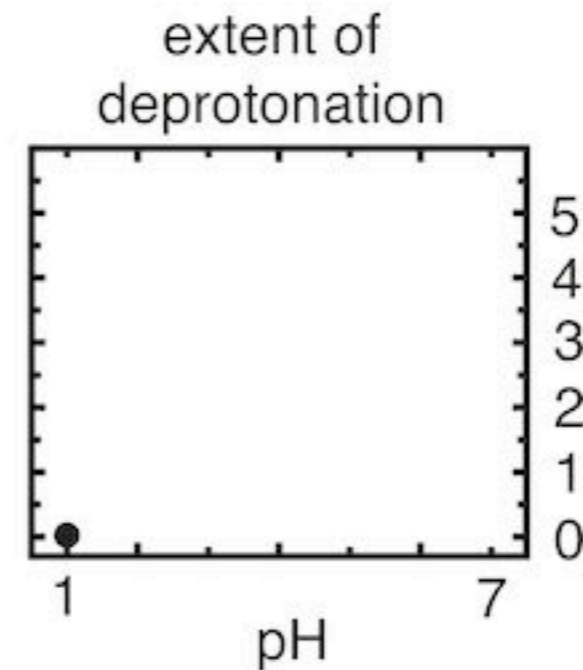
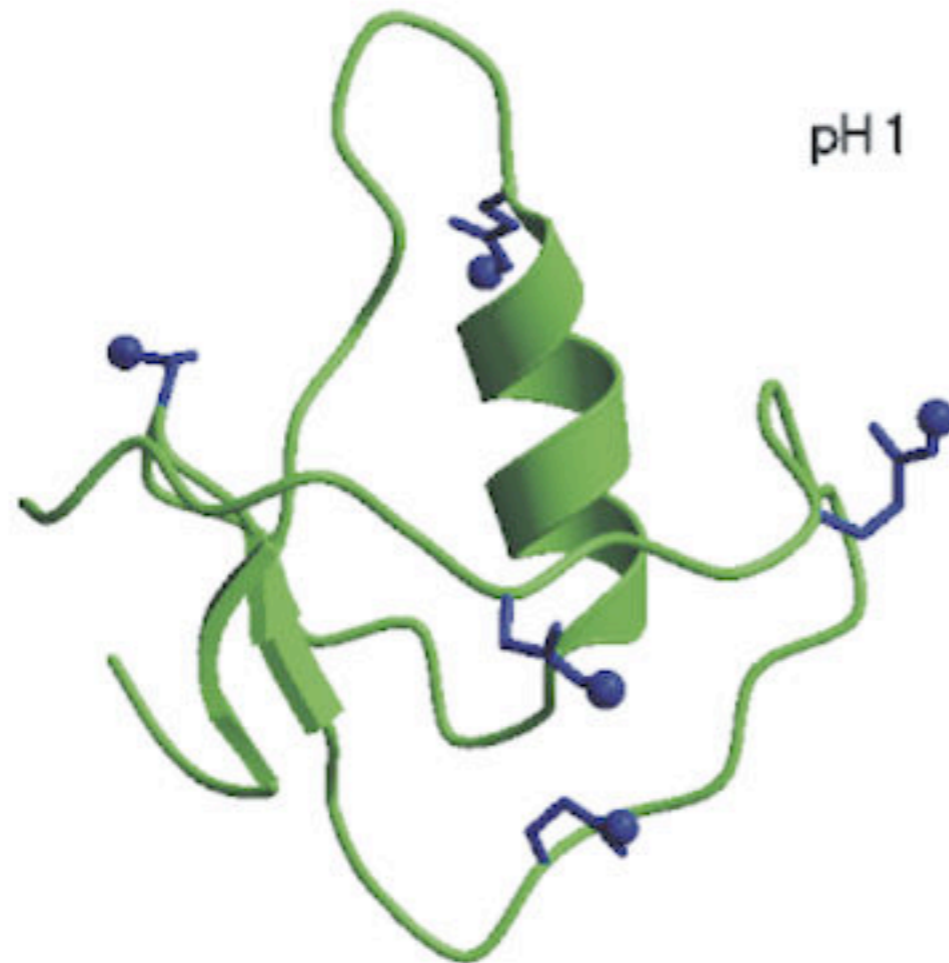
Molecular dynamics at constant pH

in silico titration experiment

MD simulations at different solvent pH values

Henderson-Hasselbalch

$$\frac{[A^-]}{[A^-] + [AH]} = \frac{1}{10^{n(\text{pK}_a - \text{pH})} + 1}$$

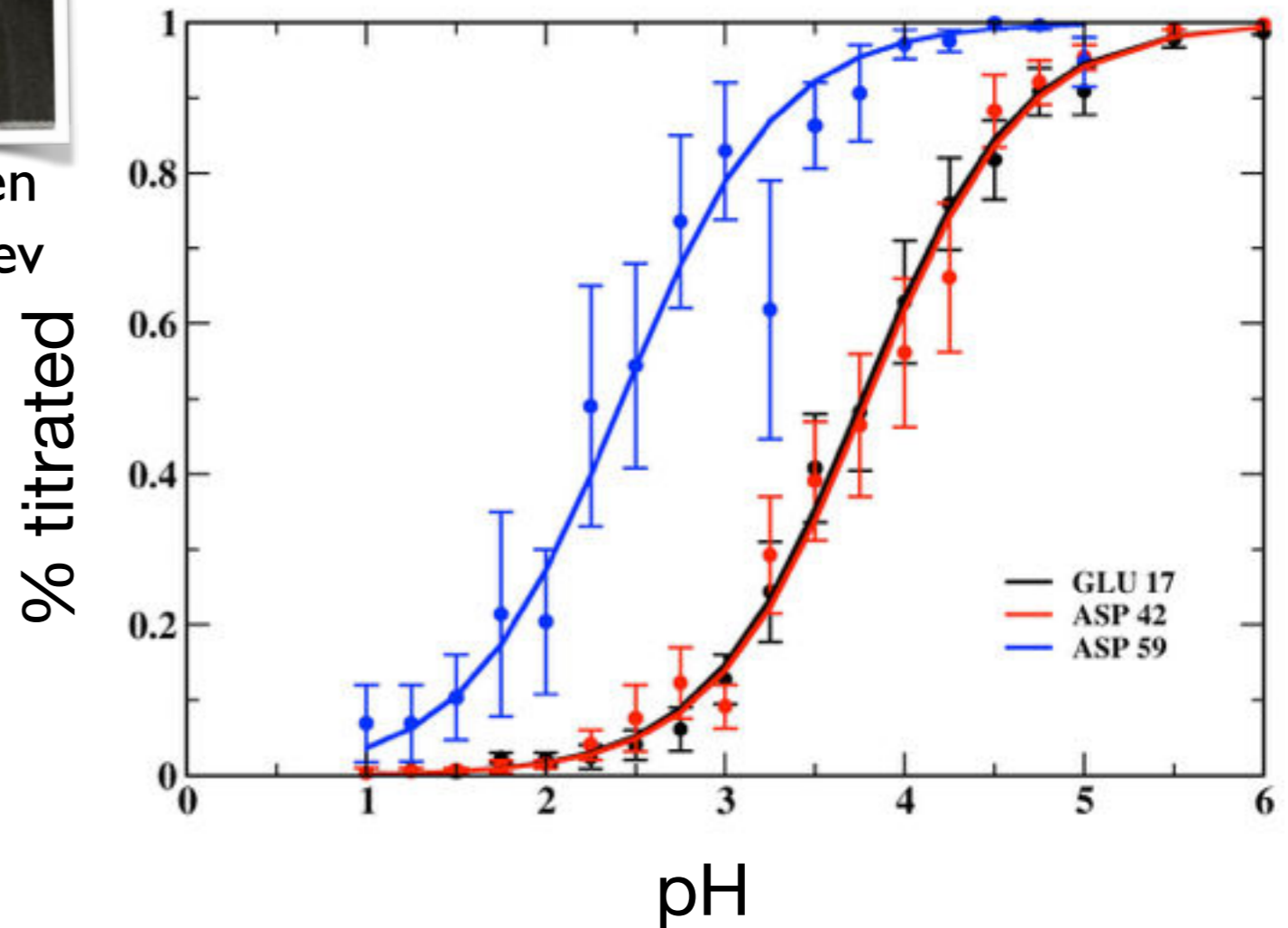
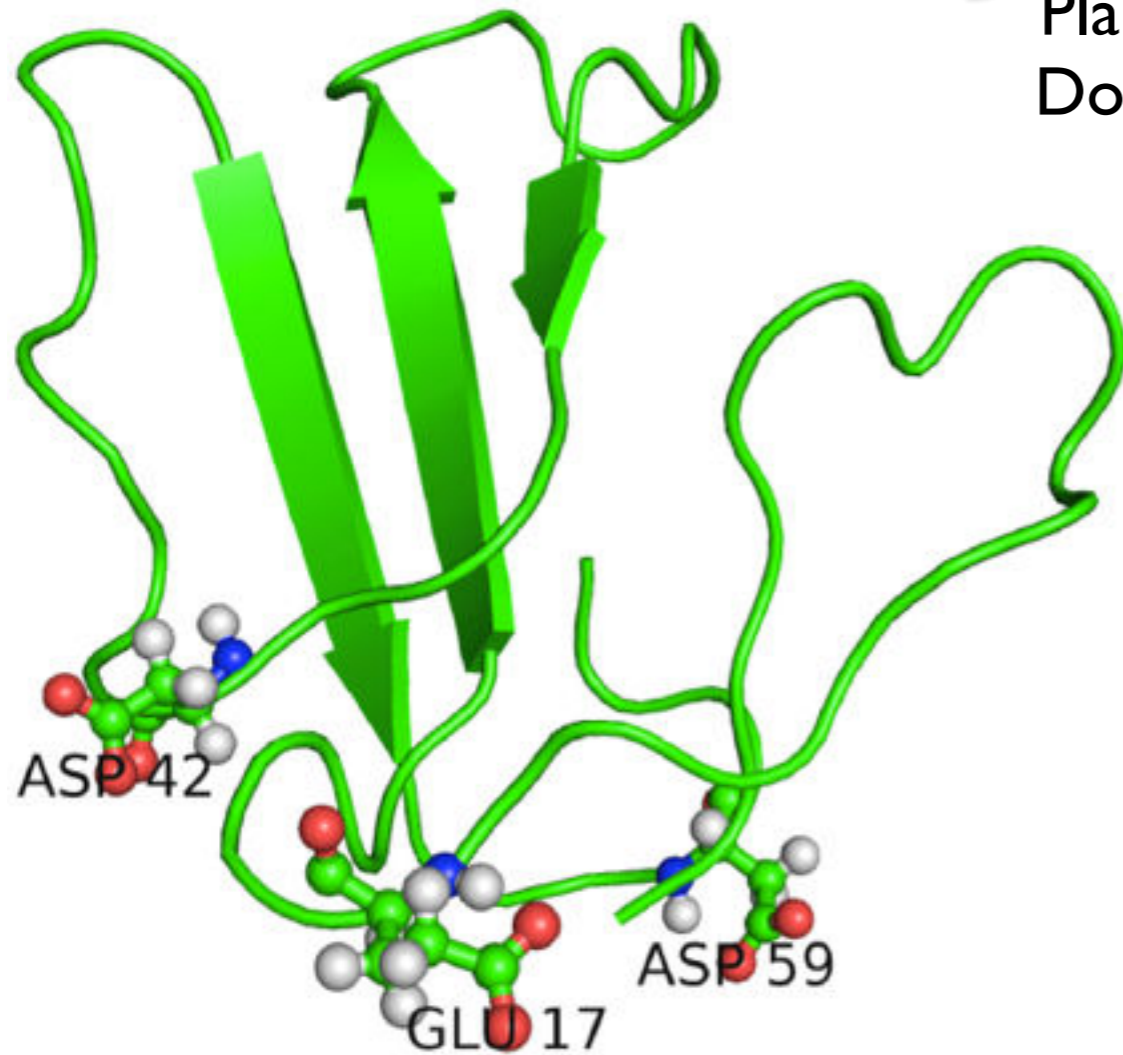


pK_a's in proteins

cardiotoxin V



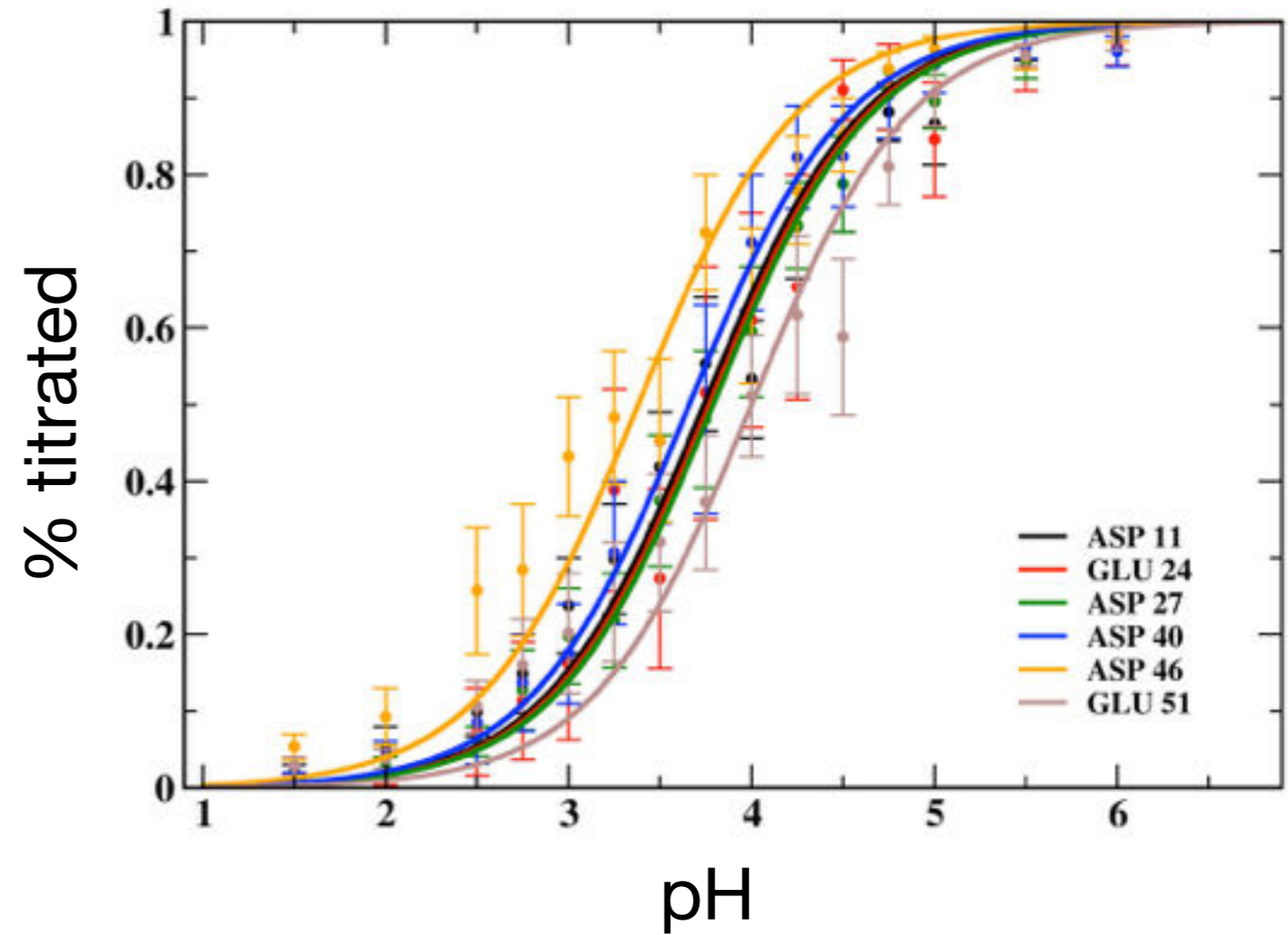
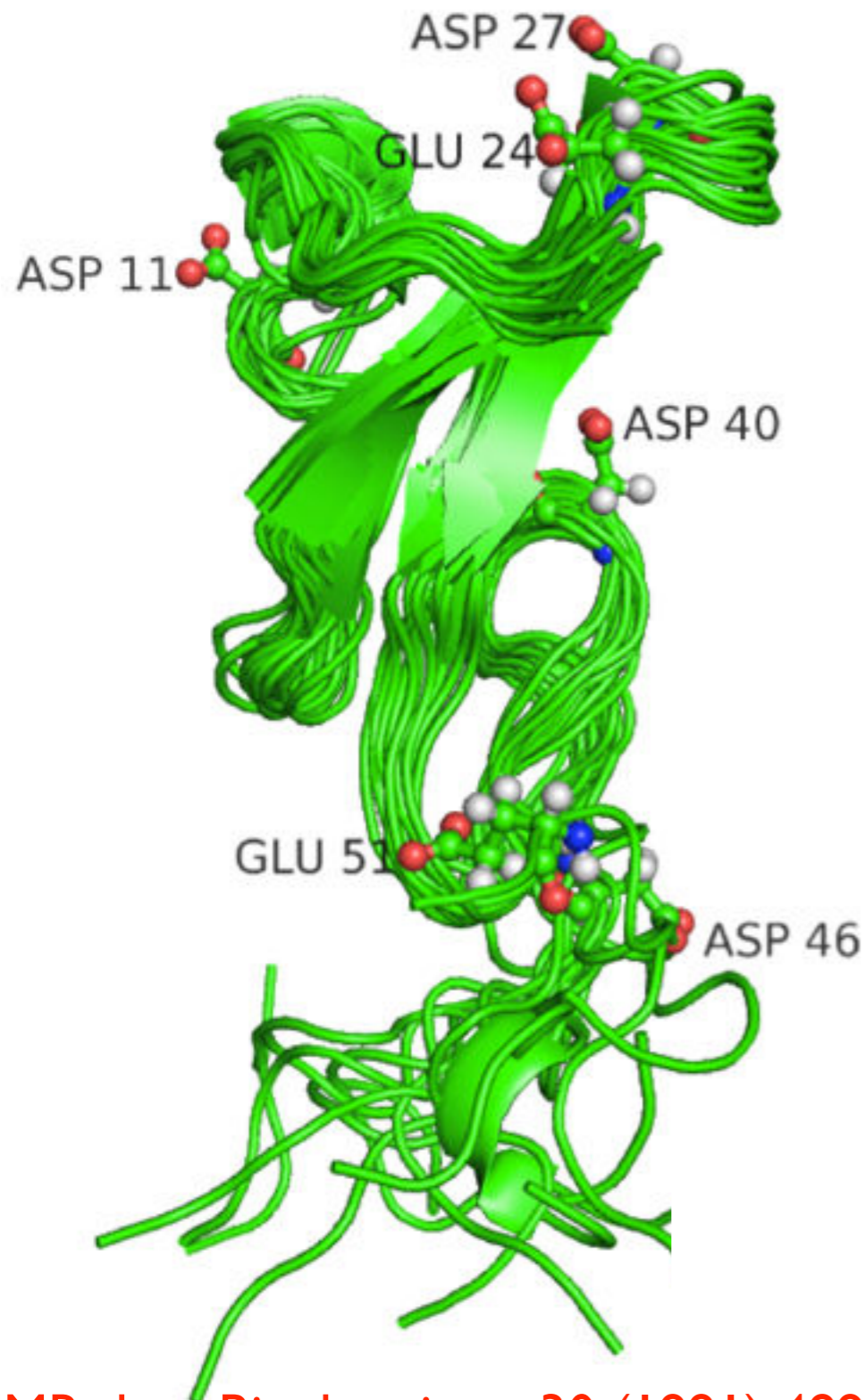
Plamen
Dobrev



residue	CPHMD	PB	NMR
Glu17	3.77 (0.06)	3,73	4
Asp42	3.8 (0.07)	3,64	3,2
Asp59	2.44 (0.12)	2,84	< 2.3

pK_a's in proteins

epidermal growth factor



residue	CPHMD	PB	NMR
Asp11	3.74 (0.07)	4.2 (0.13)	3,9
Glu24	3.77 (0.12)	3.78 (0.15)	4,1
Asp27	3.79 (0.07)	3.8 (0.08)	4
Asp40	3.66 (0.09)	5.33 (0.28)	3,6
Asp46	3.38 (0.09)	4.1 (0.34)	3,8
Glu51	4.0 (0.09)	4.1 (0.24)	4

NMR data: Biochemistry 30 (1991) 4896

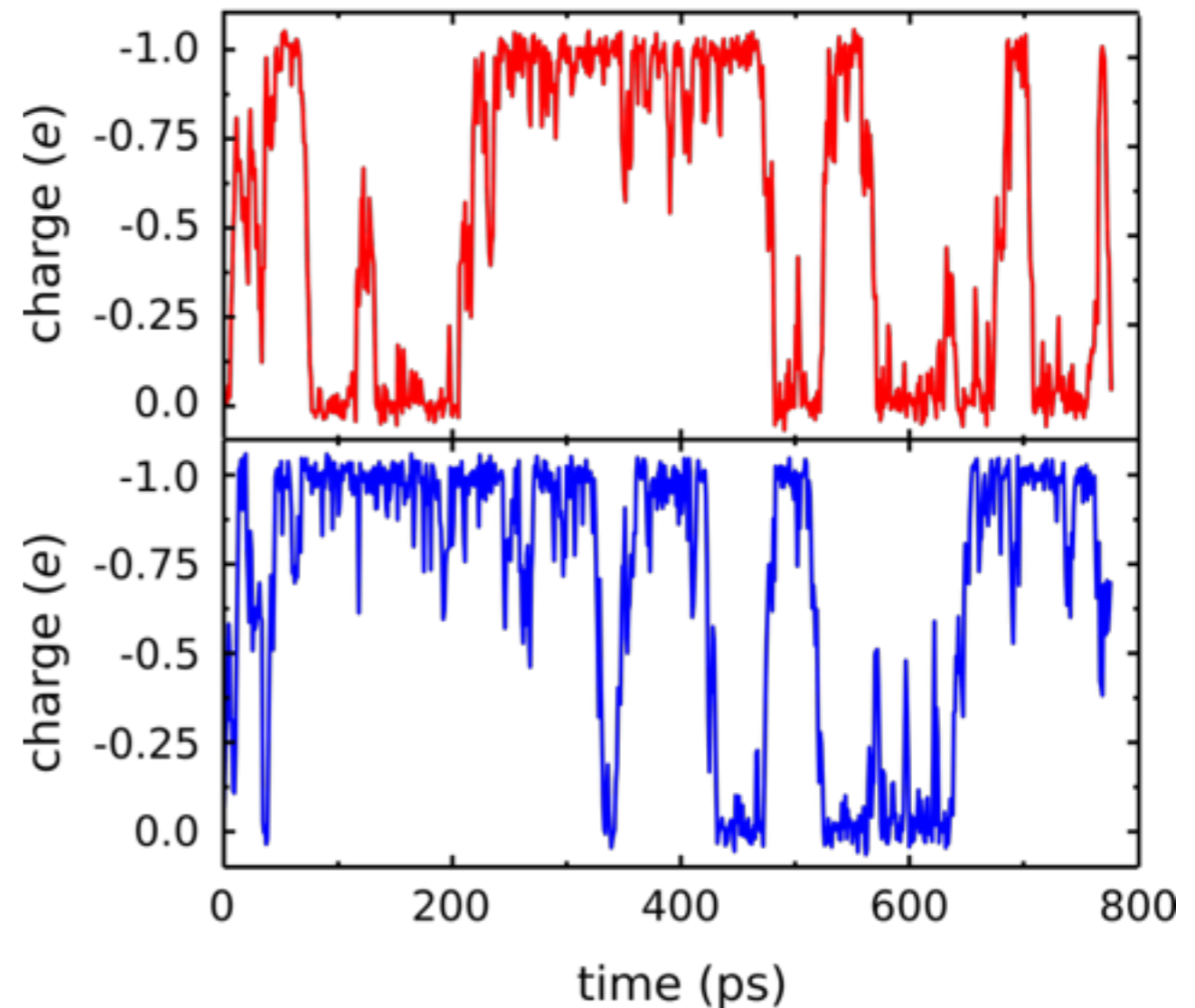
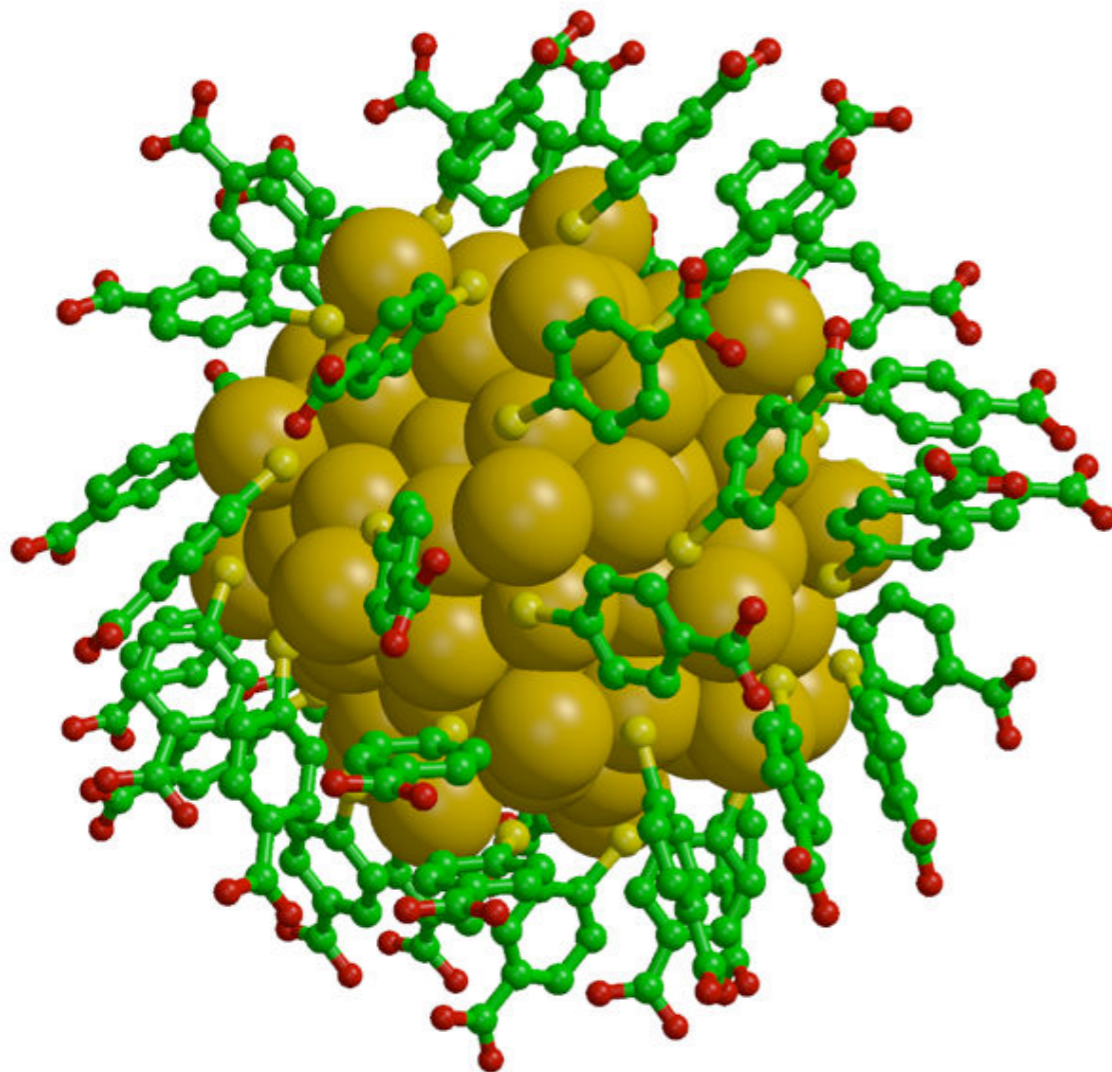
Analysis of constant pH MD of Au particles

charge fluctuations

many protonations & deprotonation events

44 titratable sites

trajectory for sites 9 & 10

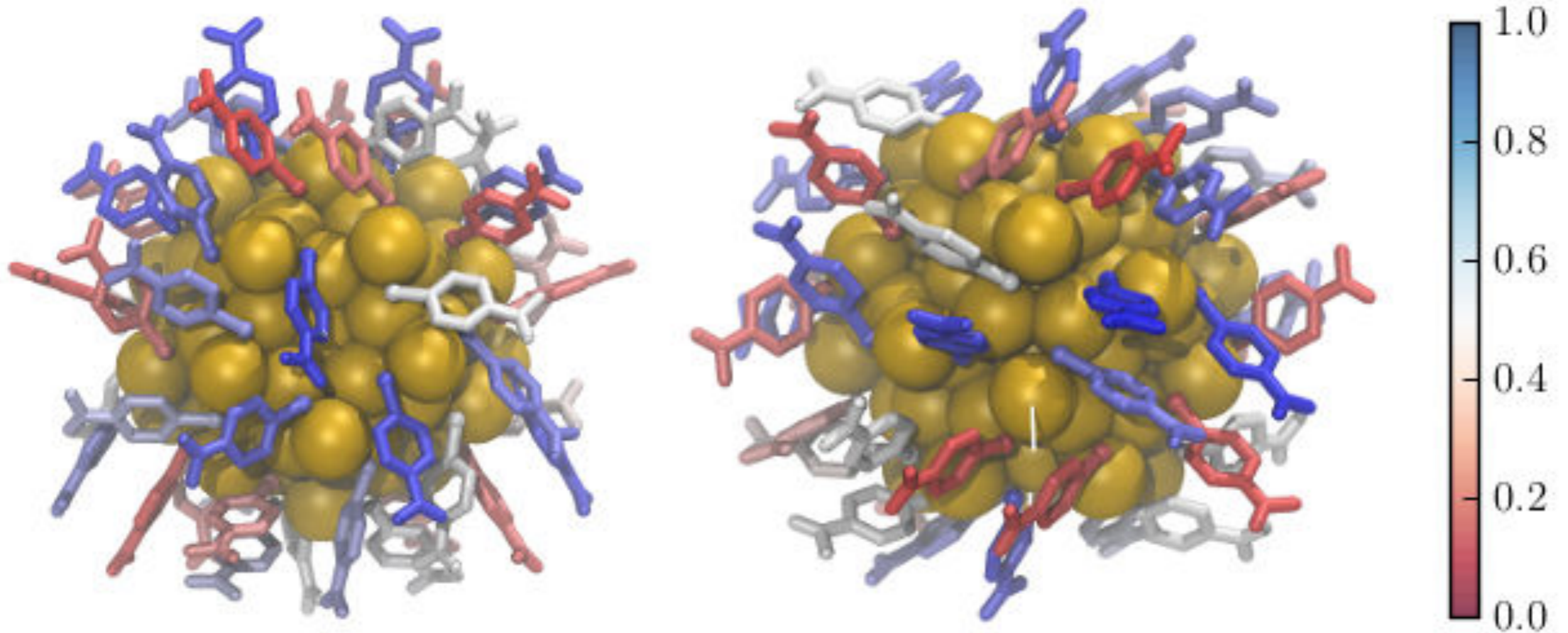


protonation of nanoparticles

are there patterns

important for interactions/recognition/biocompatibility

average



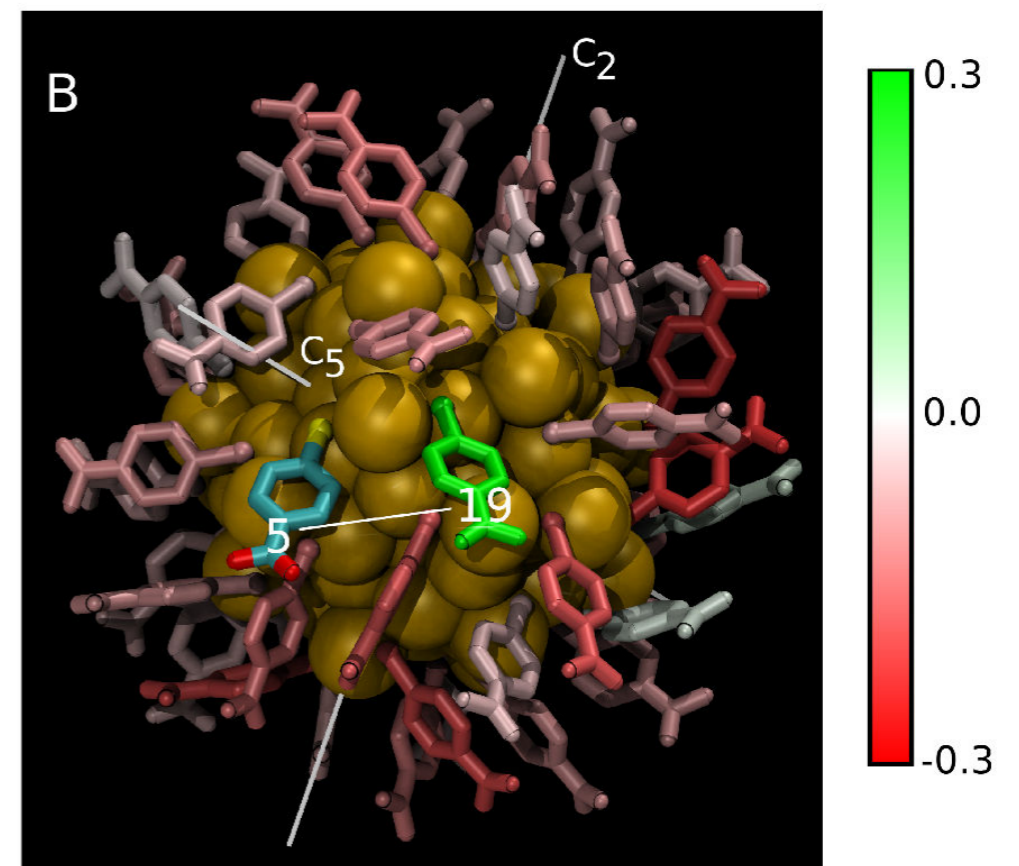
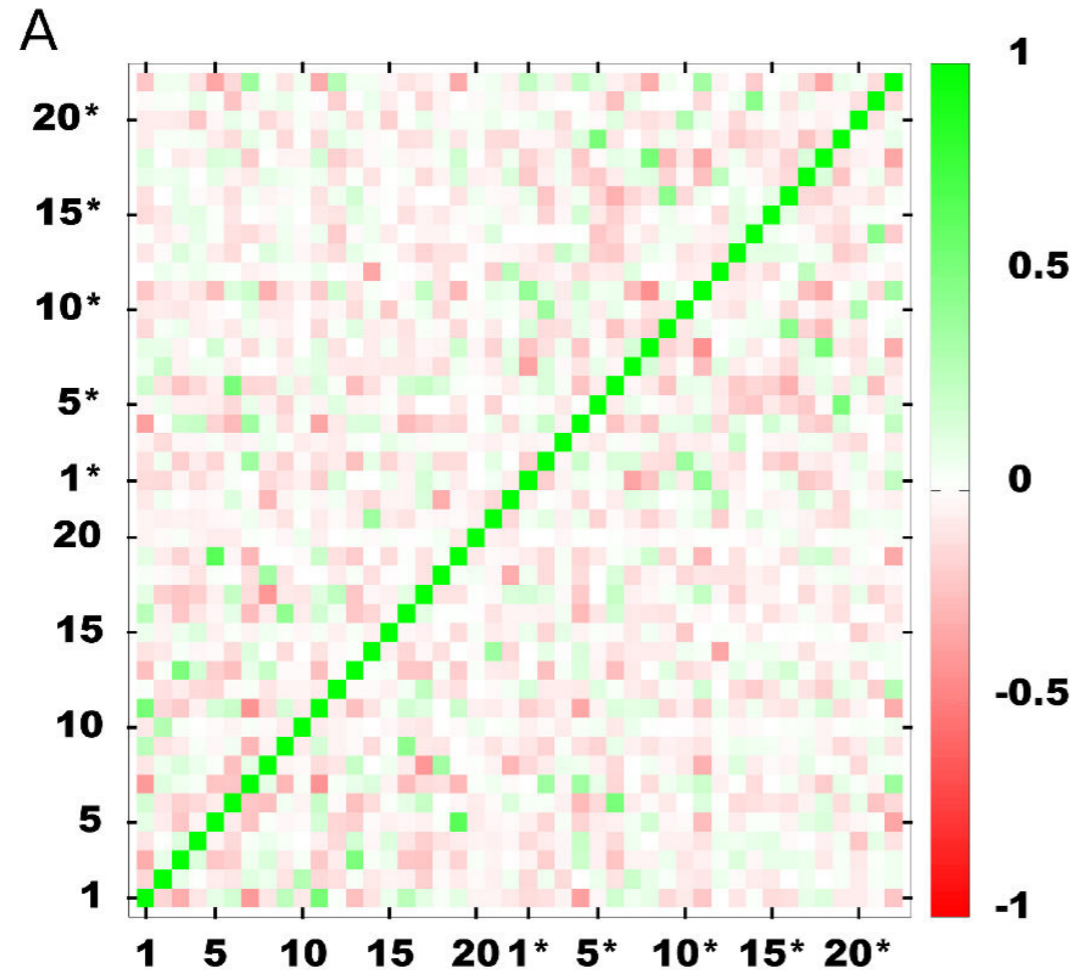
anyone sees a pattern?

protonation nanoparticles

correlations

cooperative/anti cooperative proton binding

Pearson correlation plot

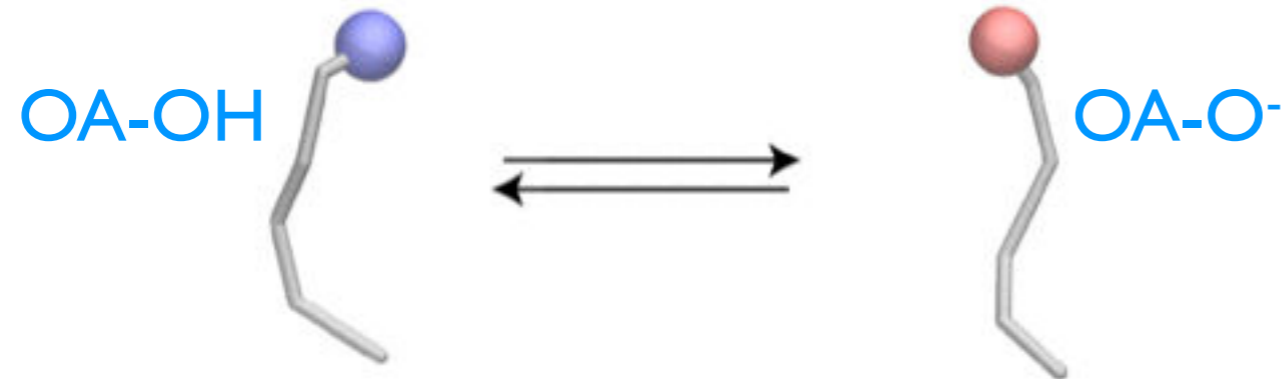


anyone sees a pattern?

Coarse grained simulations at constant pH

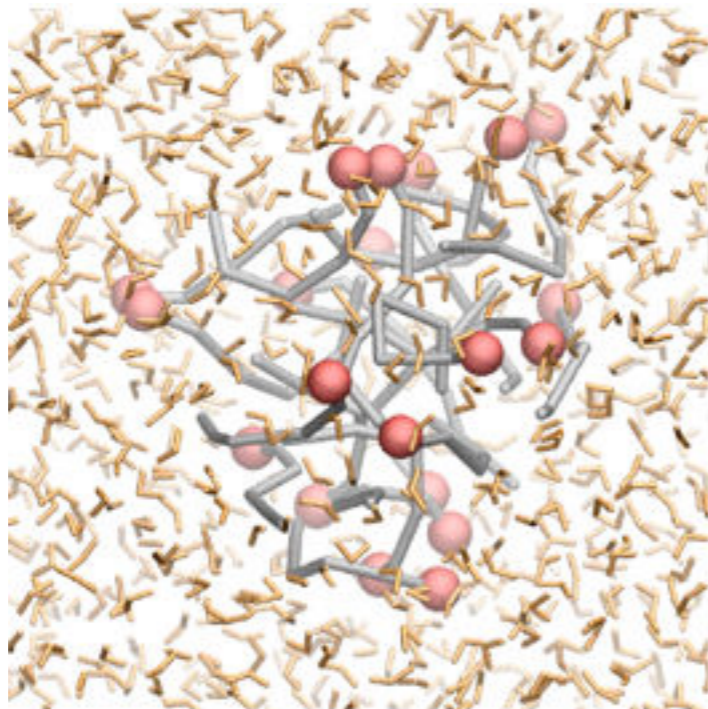
Oleic acid aggregates

MARTINI coarse grained force field (Marrink *et al.*)

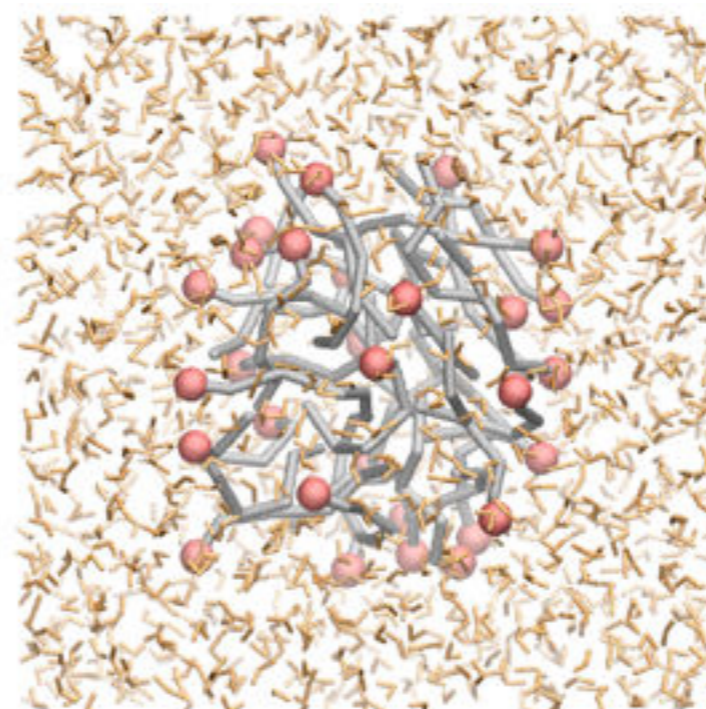


Drew
Bennett

micelle systems (self-aggregation)



20-mer



30-mer

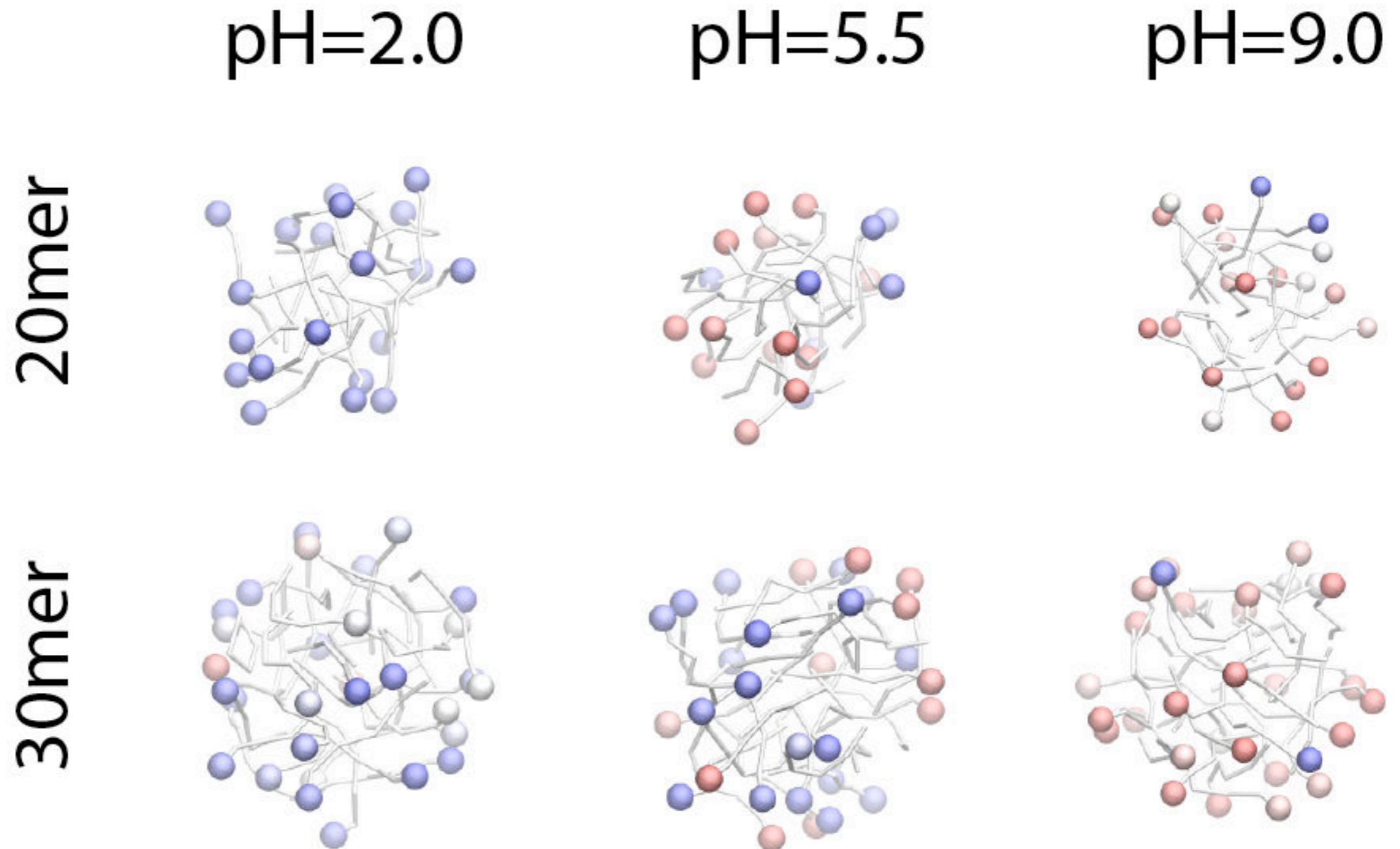


Peter
Tieleman

Coarse grained constant pH simulations

Oleic acids aggregates

micelles at different pH

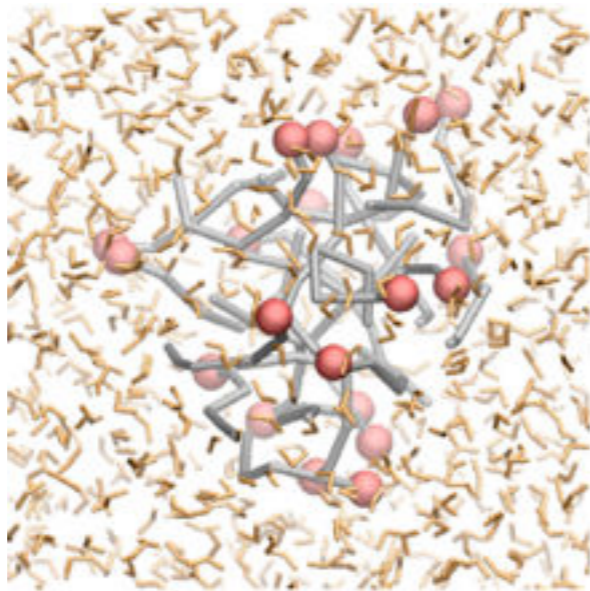


Coarse grained constant pH simulations

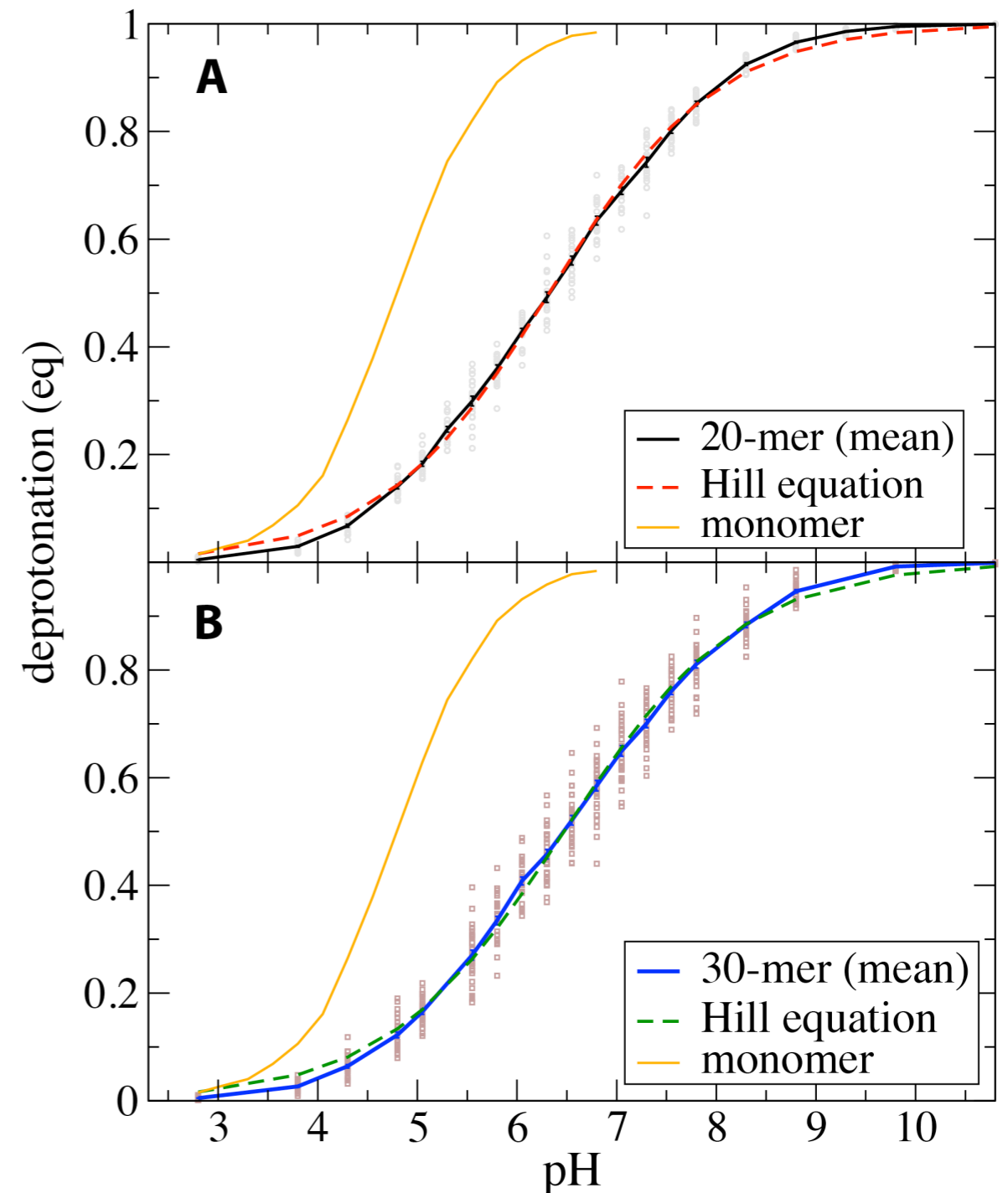
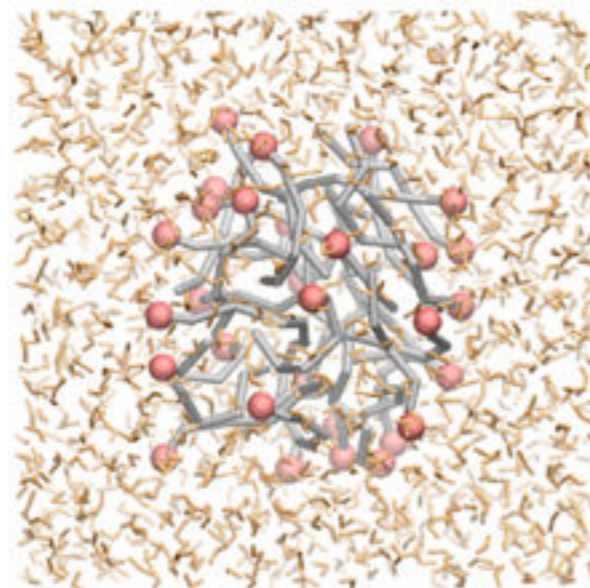
Oleic acids aggregates

upward pK_a shift of OA in micelles

20-mer



30-mer

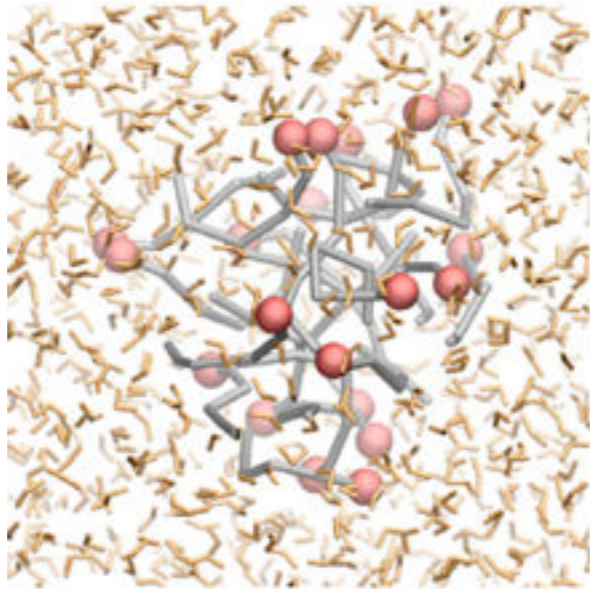


Coarse grained constant pH simulations

Oleic acids aggregates

pK_a shift correlates with micelle radius

20-mer



30-mer

